

# General Chemistry II Lyophilized Kit

### Product Name

eral Chemistry II Lyophilized Kit

# 【Packing Specification】 Type A: 1 Test / Disc, 10 Dis

Type R: 1 Test / Disc. 10 Discs / Roy

e A without diluent container; Type B with diluent contain

#### Testing Instrument

lercare M or Pointcare M chemistry analyzer

#### Intended Use

e General Ch mistry II Lyophilized Kit used with the Celercare M or the Pointcare M chemistry analyzer, is intended to be used for the in vitro quantitative determination of potassium (K<sup>+</sup>), sodium (Na<sup>+</sup>), chloride (CL<sup>-</sup>), carbon dioxide (CO<sub>2</sub>), glucose (GLU), creatinine (CRE), blood urea and amylase (AMY) in heparinized whole blood, heparinized plasma, or serum in a clinical laboratory setting or

point-of-care location.
The General Chemistry II Lyophilized Kit measurements are used in the diagnosis of salt metabolism disorders, urinary system diseases ncreas dis

## [Principles of Testing]

The General Chemistry II Lyophilized Kit is used to quantitatively test the concentration of the eight biochemical indicators in the sample, which is based on the spectrophotometry. The principles are as follows:

# Potassium (K+)

In the coupled enzyme reaction, pyruvate kinase (PK) dephosphorylates phosphoenolpyruvate (PEP) to form pyruvate.

Lactate dehydrogenase (LDH) catalyzes conversion of pyruvate to lactate, Concomitantly, NADH is oxidized to NAD+. The rate of change in absorbance due to the conversion of NADH to NAD+ is directly proportional to the amount of notassium in the sample rences from other ions are minimized with the addition of some special ingredients.  $ADP + PEP \xrightarrow{K^*, PK} PYTUVATE + ATP$   $PYTUVATE + NADH + H^+ \xrightarrow{LDH} Lactate + NADH$ 

# odium (Na+)

In the enzymatic reaction, β-D-galactosidase is activated by the sodium in the sample. The activated enzyme catalyzes the reaction of o-nitrophenyl-β-D-galactopyranoside (ONPG) to o-nitrophenol and galactose. · Na+, β-D-galactosidase

ONPG -

#### Chloride (CL-)

The method is based on the determination of chloride-dependen activation of α-amylase activity. Deactivated α-amylase is reactivated by addition of the chloride ion. The reactivation of α-amylase activity is proportional to the concentration of chloride ion in the sample. The activated α-amylase converts the substrate,

2-chloro-4-nitrophenyl-β-1,4-galactopyranosylmaltoside (CNP-G2) to

2-chloro-4-nitrophenol (CNP) producing color and 1,4-galactopyranosylmaltoside. The reaction is measured

bichromatically and the increase in absorbance is directly proportional to the reactivated α-amylase activity and the concentration of chloric ion in the sample.  $CNP-G2 \xrightarrow{Cl^-, \alpha-amylase} CNP+G2$ 

# Carbon Dioxide (CO<sub>2</sub>)

In the enzymatic method, the specimen is first made alkaline to convert all forms of carbon dioxide (CO2) to bicarbonate (HCO2) Phosphoenolpyruvate (PEP) and HCO<sub>3</sub><sup>-</sup> then react to form oxaloacetate and phosphate in the presence of phosphoenolpyruvate carboxylase (PEPC). Malate dehydrogenase (MDH) catalyzes the reaction of oxaloacetate and reduced nicotinamide adenine dinucleotide (NADH) to NAD+ and malate. The rate of change in absorbance due to the conversion of NADH to NAD+ is directly proportional to the amount of  $CO_2$  in the sample. PEP +  $HCO_3^-$  PEPC Oxaloacetate + Phosphate Oxaloacetate +  $NADH + H^+$  MDH  $NAD^+ + Malate$ 

# Glucose (GLU)

The reaction of glucose with adenosine triphosphate (ATP) catalyzed by hexokinase (HK), produces glucose-6-phosphate (G-6-P) and adenosine diphosphate (ADP). Glucose-6-phosphate dehydrogena (G-6-PDH) catalyzes the reaction of G-6-P into 6-phosphoglu and the reduction of nicotinamide adenine dinucleotide phosphate (NADP+) to NADPH.

G-6-P+NADP — G-6-PDH —, 6-Phosphate + ADP —, 6-Phosphogluconate + NADPH+H\*

The absorbance is measured bichromatically at 340 nm and 405 nm. The production of NADPH is directly proportional to the amount of glucose present in the sample.

Creatinine (CRE)

In the coupled enzyme reactions, creatinineamidohydrolase (CAH) hydrolyzes creatinine to creatine. A second enzyme, creatineamidinohydrolase (CRH), catalyzes the formation of sarcosin from creatine. Sarcosine oxidase (SAO) causes the oxidation of

sarcosine to glycine, formaldehyde and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>). In a Trinder finish, peroxidase (POD) catalyzes the reaction among the

hydrogen peroxide, 2, 4, 6-tribromo-3-hydroxybenzoic acid (TBHBA) and 4-aminoantipyrine (4-AAP) into a red quinoneimine dye.

Potassium ferrocyanide and ascorbate oxidase are added to the reaction mixture to minimize the potential interference of bilirubin at reaction mixture to minimum assorbic acid respectively. 
Creatinine  $+ H_2O \xrightarrow{CAH} \rightarrow Creatine$ Creatine  $+ H_3O \xrightarrow{CAH} \rightarrow Creatine$ Creatine  $+ H_3O \xrightarrow{CAH} \rightarrow Creatine$   $+ H_2O + O_2 \xrightarrow{SAO} \rightarrow Glycine + Formaldehyde + H_2O_2$   $+ H_2O + O_3 \xrightarrow{RO} \rightarrow Glycine + Formaldehyde + H_3O_3$ Red Quinoneimine Dye  $+ H_3O_3$ 

H<sub>2</sub>O<sub>2</sub> +TBHBA + 4-AAP POD Red Quinoneimine Dye + H<sub>2</sub>O Two cuvettes are used to determine the concentration of creatinine in

the sample. Endogenous creatine is measured in the blank cuvette, which is subtracted from the combined endogenous creatine and the reatine formed from the enzyme reactions in the test cuvette. Once

the endogenous creatine is eliminated from the calculations, the concentration of creatinine is proportional to the intensity of the red color produced. The endpoint reaction is measured as the difference in ance at 546 nm and 700 nm.

# Urea

In the coupled-enzyme reaction, urease hydrolyzes urea into amn and carbon dioxide. Upon combining ammonia withα-oxoglutarate and reduced nicotinamide adenine dinucleotide (NADH), the enzyme glutamate dehydrogenase (GLDH) oxidizes NADH to NAD+.

Urea + 2H<sub>2</sub>O Urease 2NH<sub>4</sub>+ + CO<sub>2</sub>2

NAD+

The rate of change of the absorbance difference between 340 nm and 405 nm is caused by the conversion of NADH to NAD<sup>+</sup> and is directly proportional to the amount of urea present in the sample.

### Amylase (AMY)

zyme reaction, amylase in the sample hydrolyze 2-chloro-4-nitrophenyl-β-1,4-galactopyranosylmaltoside (CNP-G2) to

2-chloro-4-nitrophenol (CNP) producing color and

1.4-galactopyranosylmaltoside. The change in absorbance of the CNP onal to the amylase activity in the sample at 405 n ctly proportion and505 nm.

CNPG2 \_\_AWY ... CNP + G2

# Principle of Operation

Refer to the Celercare M or the Pointcare M chemistry analyzer Operator's Manual, for the Principles and Limitations of the Procedure

# Description of Reager

follows (after redissolution):

Each General Chemistry II Lyophilized Kit contains lyophilized test-specific reagent beads. A lyophilized blank reagent in each disc for a judgment of error 0209.

Type B is the reagent disc with diluent container

Type A is the reagent disc without diluent container

Stabilizer

Calibration information is included in barcode code. Please check it on the label. ent of each General Chemistry II Lyophilized Kit is as The comp

Component Quantity Potassium assay reagent 13.5 μL Sodium assav reagent 13.5 uL Chloride assay reagent 13.5 μL Carbon dioxide assay reagent 5.3 μL Glucose assay reagent Creatinine assay reagent 6.6 µL 3.5 μL Urea assay reagent 13.5 uL Amylase assay reagent 13.5 μL Appropriate

# [Storage]

Store reagent discs in their sealed pouches at 2-8°C (36-46°F). Do not expose opened or unopened discs to direct sunlight or temperatur above 32°C (90°F). Reagent discs may be used until the expiration date included on the package. The expiration date is also encoded in the unique code printed on the sealing pouch. An error message will appear on the Celercare M or the Pointcare M chemistry analyzer isplay if the reagents have expired

A torn or otherwise damaged pouch may allow moisture to reach the unused disc and adversely affect reagent performance. Do not use a disc from a damaged pouch

【Sample Requirements】
Sample collection techniques are described in the "Sample requirement" section of the Celercare M or the Pointcare M chemistry analyzer Operator's Manual.

The required sample usage is 100 μL of lithium heparin whole blood, lithium heparin plasma, serum or quality controls. Pla when using Type A. The required diluent usage is 430  $\mu L$  of sterilized water for injection.

Whole blood samples collected by venipuncture must be homogeneous efore transferring the sample to a reagent disc

At the same time, it is necessary to carry out the test within 60 minutes Before taking the test, shake the lithium heparin blood collection tube gently upside down several times.

The glucose concentration is affected by the patient's feeding time and the storage environment after the sample is collected. In order to accurately measure glucose, a sample of the patient should be taken after at least 12 hours of fasting. For uncentrifuged samples stored a room temperature, the glucose concentration is reduced by about 5-12 Use only lithium heparin evacuated specimen collection tubes for

whole blood or plasma samples.

The test was started within 10 minutes after transferring the sample to

# the reagent disc 【Interfering Substances】

Studies on known drugs or chemicals have found that when the interfering substances contained in the sample exceed the contents in the table below, the final test results are affected.

Interfering substances concentration  $(\leq)$ Bilirubin Intralipid Hemoglobin Vitamin C Creatine Analyte chloride mg/dL mmol/L 150 50 K+ Na 10 150 50 75 Cl-CO<sub>2</sub> 45 525 250 75 GLU 600 1000 40 600 CRE 40 1050 500 25 IIRFA 25 600 1000 AMY 40 1000 400

### Procedure Materials Provided

# General Chemistry II Lyophilized Kit

Celercare M or Pointcare M chemistry analyzer

Please add diluent into the diluent port when using Type A (sterilized water for injection); please tear off the aluminum strip before using for Type B.

Transfer pipettes (fixed volume 100 μL for sample and 430 μL for

### diluent) and tips Test Procedure

The complete sample collection and step-by-step operating procedures are detailed in the Celercare M or the Pointcare M chemistry analyzer

# Operator's Manual.

Each batch of reagent is calibrated using Rondox standard serum to obtain the disc-specific calibration parameters before shipment. The calibration parameters stored in the two-dimentional code printed on the sealed pouch are provided to analyzer at the time of sca

Refer to the Celercare M or the Pointcare M chemistry Operator's Manual for the specific information

# Quality Control

the code.

Refer to Operator's Manual of the Celercare M or the Pointcare M

M chemistry analyzer can be verified by running controls. For a list of approved quality control materials with acceptance ranges.

If control results are out of range, repeat one time. If still out of range, call MNCHIP customer service or local distributers for technical support. Do not report the results if controls are outside their labeled

# Results

Manual.

The Celercare M or the Pointcare M chemistry analyzer automatic calculates and prints the analyte concentrations in the sample. Deta of the endpoint and rate reaction calculations are found in the re M or the Pointcare M chemistry analyzer Oper

These ranges are provided as a guideline only. It is recommended that your office or institution establish normal ranges for your particular

Analyte	SI Units	Common Units
K <sup>+</sup>	Serum: 3.5 ~ 5.3 mmol/L	Serum: 3.5 ~ 5.3 mmol/L
	Whole blood and plasma:	Whole blood and plasma:
	$3.0 \sim 5.1 \text{ mmol/L}$	$3.0\sim5.1\ mmol/L$
Na <sup>+</sup>	137 ~ 147 mmol/L	137 ~ 147 mmol/L
Cl·	99 ~ 110 mmol/L	99 ~ 110 mmol/L
$CO_2$	23 ~ 29 mmol/L	23 ~ 29 mmol/L
GLU	3.9 ~ 6.1 mmol/L	70.2 ~ 109.8 mg/dL
CRE	Male: 54 ~ 109 μmol/L;	Male: 0.61 ~ 1.23 mg/dL;
	Female: 45 ~ 84 µmol/L	Female: 0.51~ 0.95 mg/dL
UREA	2.9 ~ 8.2 mmol/L	17.42 ~ 49.25 mg/dL
AMY	0 ~ 220 U/L	0 ~ 220 U/L

# [Interpretation of Results]

Physiological interferents (hemolysis, icterus and lipemia) cause changes in the reported concentrations of some analytes. The sample indices are printed on the bottom of each printout to inform the operator about the abnormal sample. The operator should avoid sample hemolysis caused by irregular blood collection.

The Celercare M or the Pointcare M chemistry analyzer suppresses any results that are affected by >10% interference from hemolysis lipemia or icterus. "HEM", "LIP", or "ICT" respectively, is printed on the printout in place of the result.

Any result for a particular test that exceeds the assay range should be analyzed by another approved test method or sent to a referral laboratory. Do not dilute the sample and run it again on the Celercare M or the Pointcare M chemistry analyzer.

### Limitations of Procedure

The General Chemistry II Lyophilized Kit should be used with the Celercare M or the Pointcare M chemistry analyzer, and is just used for in vitro diagnosis (IVD).

As with any diagnostic test procedure, all other test procedures including the clinical status of the patient, should be considered prior al diagnosis

### Performance Characteristics

Analyte	The relative deviation or absolute deviation should meet the following requirements
K+	B% ≤ 15.0%
Na <sup>+</sup>	B% ≤ 15.0%
Cl-	B% ≤ 15.0%
$CO_2$	$B\% \le 10.0\%$
GLU	$B\% \le 20.0\%$
CRE	$B\% \le 10.0\%$
UREA	$B\% \le 15.0\%$
AMY	$B\% \le 10.0\%$

Satch precision		
Analyte	Coefficient of variation (≤*)	
K+	5.0%	
Na <sup>+</sup>	5.0%	
Cl-	5.0%	
$CO_2$	5.0%	
GLU	5.0%	
CRE	5.0%	
UREA	5.0%	
$\Delta MY$	5.0%	

# Inter batch precision

inci batch precision		
Analyte	Relative Range (≤ *)	
K <sup>+</sup>	10.0%	
Na <sup>+</sup>	10.0%	
Cl-	10.0%	
$CO_2$	10.0%	
GLU	10.0%	
CRE	10.0%	
UREA	10.0%	
AMY	10.0%	

Dynamic Ranges		
Analyte	Dynamic Ranges	
K <sup>+</sup>	1 ~ 8 mmol/L	
Na <sup>+</sup>	90 ~ 170 mmol/L	
Cl-	60 ~ 140 mmol/L	
$CO_2$	10 ~ 35 mmol/L	
GLU	$1 \sim 30 \text{ mmol/L}$	
CRE	20 ~ 1500 μmol/L	
UREA	0.9 ~ 35.7 mmol/L	
AMY	$5 \sim 1100 \text{ U/L}$	

# Notes 1

ed reagent discs contain human body fluids. Follow good laboratory safety practices when handling and disposing of used discs. See the Celercare M or the Pointcare M chemistry analyzer Operator's Manual for instructions on cleaning biohazardous spills.

The reagent discs are plastic and may crack or chip if dropped. Never use a dropped disc as it may spray biohazardous material throughout or of the analyzer.

Reagent beads may contain acids or caustic substances. The operato does not come into contact with the reagent beads when following the recommended procedures. The operator should avoid ingestion, skin contact, or inhalation of the reagent beads.

The diluent can be selected from purified water having a conductivity (measured at 25°C) greater than 10 MΩ/cm, we recommend using the sterilized water for injection to reduce discrepancies or errors in test results due to the water, and it should be prevented from being

ed to the air for a long time after opening.

# [Symbols Used in Labelling]

Symbols Used in Labelling 2		
Symbol	Explanation	
IVD	In vitro diagnostic medical device	
	Manufacturer	
EC REP	Authorized representative in the European Community	
₽	Use-by date	
LOT	Batch code	
س	Date of manufacture	
Œ	CE MARK	
(II	Consult instructions for use	
zc. K	Limit of temperature	
UDI	Unique device identifier	
(2)	Do not re-use	

# [Manufacturer]

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