

[Product Name]

Equine Profile (15+4)

[Packing Specification]

Type A: 1 Test / Disc, 10 Discs / Box;

Type B: 1 Test / Disc, 10 Discs / Box.

Type A without diluent container; Type B with diluent container.

Testing Instrument

Celercare V or Pointcare V chemistry analyzer

[Intended Use]

The Equine Profile (15+4) used with the Celercare V or the Pointcare V chemistry analyzer, is intended to be used for the in vitro quantitative determination of total Protein (TP), albumin (ALB), total bilirubin (TBIL), creatinine (CRE), creatine kinase (CK), glucose (GLU), lactate dehydrogenase (LDH), potassium (K⁺), sodium (Na⁺), calcium (Ca), aspartate aminotransferase (AST), gamma glutamyltransferase (GGT), total carbon dioxide (tCO₂), urea nitrogen(BUN) and alkaline phosphatase (ALP) in heparinized plasma, or serum in a clinical laboratory setting or point-of-care location.

The Equine Profile (15+4) measurements are used in the diagnosis of liver and gallbladder diseases, glucose metabolism and lipid metabolism disorders, water and salt metabolism disorder, pancreatic diseases, cardiovascular diseases, urinary system diseases.

(Principles of Testing)

The Equine Profile (15+4) is used to quantitatively test the concentration of the 15 biochemical indicators in the sample, which is based on the spectrophotometry. The principles are as follows:

1. Total Protein (TP)

The total protein method is a Biuret reaction, the protein solution is treated with cupric [Cu(II)] ions in a strong alkaline medium. The Cu(II) ions react with peptide bonds between the carbonyl oxygen and amide nitrogen atoms to form a colored Cu-protein complex.

The amount of total protein present in the sample is directly proportional to the absorbance of the Cuprotein complex. The total protein test is an endpoint reaction and the absorbance is measured as the difference in absorbance between 546 nm and 800 nm.

2. Albumin (ALB)

Bromcresol green (BCG), when bound with albumin, changes color from a yellow to green color. The absorbance maximum changes with the color shift.

Bound albumin is proportional to the concentration of albumin in the sample. This is an endpoint reaction that is measured as the difference in absorbance between 600 nm and 700 nm.

3. Total Bilirubin (TBIL)



In the enzyme procedure, bilirubin is oxidized by bilirubin oxidase (BOD) into biliverdin. Bilirubin is quantitated as the difference in absorbance between 450nm and 546 nm. The initial absorbance of this endpoint reaction is determined from the bilirubin blank cuvette and the final absorbance is obtained from the bilirubin test cuvette. The amount of bilirubin in the sample is proportional to the difference between the initial and final absorbance measurements.

Bilirubin +
$$O_2 \longrightarrow Biliverdin + H_2O$$

4. Urea Nitrogen (BUN)

In the coupled-enzyme reaction, urease hydrolyzes urea into ammonia and carbon dioxide. Upon combining ammonia withα-oxoglutarate and reduced nicotinamide adenine dinucleotide (NADH), the enzyme glutamate dehydrogenase (GLDH) oxidizes NADH to NAD⁺.

$$Urea + 2H_2O \xrightarrow{Urease} 2NH_4{}^+ + CO_3{}^{2-}$$

$$NH_4{}^+ + \alpha\text{-}Oxoglutarate} + NADH \xrightarrow{\text{Q.DH}} L\text{-}Glutamate} + H_2O + NAD^+$$

The rate of change of the absorbance difference between 340 nm and 405 nm is caused by the conversion of NADH to NAD+ and is directly proportional to the amount of urea present in the sample.

5. Creatinine (CRE)

In the coupled enzyme reactions, creatinineamidohydrolase (CAH) hydrolyzes creatinine to creatine. A second enzyme, creatineamidinohydrolase (CRH), catalyzes the formation of sarcosine from creatine. Sarcosine oxidase (SAO) causes the oxidation of sarcosine to glycine, formaldehyde and hydrogen peroxide (H₂O₂). In a Trinder finish, peroxidase (POD) catalyzes the reaction among the hydrogen peroxide, 2, 4, 6-tribromo-3-hydroxybenzoic acid (TBHBA) and 4-aminoantipyrine (4-AAP) into a red quinoneimine dye. Potassium ferrocyanide and ascorbate oxidase are added to the reaction mixture to minimize the potential interference of bilirubin and ascorbic acid respectively.

Creatinine +
$$H_2O$$
 \longrightarrow Creatine

Creatine + H_2O \longrightarrow Sarcosine + Urea

Sarcosine + H_2O + O_2 \longrightarrow Glycine + Formaldehyde + H_2O_2
 H_2O_2 +TBHBA + 4-AAP \longrightarrow Red Quinoneimine Dye + H_2O_2

Two cuvettes are used to determine the concentration of creatinine in the sample. Endogenous creatine is measured in the blank cuvette, which is subtracted from the combined endogenous creatine and the creatine formed from the enzyme reactions in the test cuvette. Once the endogenous creatine is eliminated from the calculations, the concentration of creatinine is proportional to the intensity of the red color produced. The endpoint reaction is measured as the difference in absorbance at 546 nm and 700 nm.

6. Glucose (GLU)

The reaction of glucose with adenosine triphosphate (ATP) catalyzed by hexokinase (HK), produces glucose-6-phosphate (G-6-P) and adenosine diphosphate (ADP). Glucose-6-phosphate dehydrogenase (G-6-PDH) catalyzes the reaction of G-6-P into 6-phosphogluconate and the reduction of nicotinamide adenine dinucleotide phosphate (NADP+) to NADPH.

$$Glucose + ATP \xrightarrow{HK} Glucose-6-Phosphate + ADP$$

$$G-6-P + NADP^{+} \xrightarrow{G-6-PDH} 6-Phosphogluconate + NADPH+H^{+}$$



The absorbance is measured bichromatically at 340 nm and 405 nm. The production of NADPH is directly proportional to the amount of glucose present in the sample.

7. Creatine Kinase (CK)

Creatine kinase catalyzes the formation of creatine and adenosine triphosphate (ATP) from creatine phosphate and adenosine diphosphate (ADP). With hexokinase (HK) as a catalyst, ATP reacts with D-glucose to form ADP and D-glucose-6-phosphate (G-6-P), which is reacted with nicotinamide adenine dinucleotide phosphate (NADP⁺) in the presence of glucose-6-phosphate dehydrogenase (G-6-PDH) to produce 6-Phosphogluconate (6-PG) and NADPH.

The formation of NADPH is measured as a change in absorbance at 340 nm relative to 405 nm. This absorbance change is directly proportional to creatine kinase activity in the sample.

Creatine phosphate + ADP
$$\xrightarrow{CK}$$
 Creatine + ATP

ATP + D-glucose \xrightarrow{HK} ADP + G-6-P

G-6-P + NADP+ $\xrightarrow{G-6-PDH}$ 6-Phosphogluconate + NADPH + H+

8. Alkaline Phosphatase (ALP)

Under the catalysis of ALP, the Phosphoric acid on nitrobenzene (4-NNP) was turned into Para nitro phenol (4-NP).4-NP shows a yellow color in alkaline solution. At the wavelength of 405/505nm, the ALP activity can be calculated by monitoring the absorbance change rate.

$$4-NNP \xrightarrow{ALP} Acyl phosphate + 4-NP$$

9. Aspartate Aminotransferase (AST)

AST catalyzes the reaction of L-aspartate and α -ketoglutarate into oxaloacetate and L-glutamate. Oxaloacetate is converted to malate and NADH is oxidized to NAD+ by the catalyst MDH.

$$L\text{-aspartate} + \alpha\text{-ketoglutarate} \xrightarrow{AST} Oxaloacetate + L\text{-glutamate}$$

$$Oxaloacetate + NADH \xrightarrow{MDH} Malate + NAD^+$$

The rate of absorbance change at 340 /405 nm caused by the conversion of NADH to NAD⁺ is directly proportional to the amount of AST present in the sample.

10. Potassium (K+)

In the coupledenzyme reaction, pyruvate kinase (PK) dephosphorylates phosphoenolpyruvate (PEP) to form pyruvate. Lactate dehydrogenase (LDH) catalyzes conversion of pyruvate to lactate. Concomitantly, NADH is oxidized to NAD⁺. The rate of change in absorbance due to the conversion of NADH to NAD⁺ is directly proportional to the amount of potassium in the sample.

Interferences from other ions are minimized with the addition of some special ingredients.

ADP + PEP
$$\xrightarrow{K^+, PK}$$
 Pyruvate + ATP

Pyruvate + NADH + H⁺ \xrightarrow{LDH} Lactate + NAD⁺

11. Sodium (Na⁺)

In the enzymatic reaction, β -D-galactosidase is activated by the sodium in the sample. The activated enzymecatalyzes the reaction of o-nitrophenyl- β -D-galactopyranoside (ONPG) to o-nitrophenolandgalactose.



$$ONPG \xrightarrow{\qquad Na^+, \quad \beta\text{-D-galactosidase}} \quad \text{o-Nitrophenol} + Galactose$$

12. Calcium (Ca)

Calcium in the patient sample binds with arsenazo III to form a calcium-dye complex.

It is an endpoint reaction. The amount of total calcium in the sample is proportional to the absorbance.

13. Lactate Dehydrogenase (LDH)

Lactate dehydrogenase (LDH) catalyzes the oxidation of L-lactate to pyruvate with the concurrent reduction of nicotinamide adenine dinucleotide (NAD⁺) to reduced nicotinamide adenine dinucleotide (NADH). The NADH is then oxidized with the simultaneous reduction of INT in a reaction catalyzed by diaphorase. The intensity of the highly colored formazan is measured bichromatically at 505/800 nm and is directly proportional to the concentration of triglycerides in the sample.

$$\begin{array}{c} \text{L-Lactate} + \text{NAD}^{+} \xrightarrow{\text{LDH}} & \text{Pyruvate} + \text{NADH} + \text{H}^{+} \\ \\ \text{NADH} + \text{H}^{+} + \text{INT} \xrightarrow{\text{Diaphorase}} & \text{NAD}^{+} + \text{Formazan} \end{array}$$

14. Total Carbon Dioxide (tCO2)

In the enzymatic method, the specimen is first made alkaline to convert all forms of carbon dioxide (CO₂) to bicarbonate (HCO₃⁻). Phosphoenolpyruvate (PEP) and HCO₃⁻ then react to form oxaloacetate and phosphate in the presence of phosphoenolpyruvate carboxylase (PEPC). Malate dehydrogenase (MDH) catalyzes the reaction of oxaloacetate and reduced nicotinamide adenine dinucleotide (NADH) to NAD⁺ and malate. The rate of change in absorbance due to the conversion of NADH to NAD⁺ is directly proportional to the amount of CO₂ in the sample.

PEP + HCO₃
$$\xrightarrow{\text{PEPC}}$$
 Oxaloacetate + Phosphate

Oxaloacetate + NADH + H⁺ $\xrightarrow{\text{MDH}}$ NAD⁺+Malate

15. Gamma Glutamyltransferase (GGT)

The addition of sample containing gammaglutamyltranferase to the substrates L- γ -glutamyl-3-carboxy-4-nitroanilide and glycylglycinecauses the formation of L- γ -glutamyl-glycylglycine(glu-gly-gly) and 5-Amino-2-nitrobenzoate.

$$L$$
- γ -glutamyl-3-carboxy-4-nitroanilide+ glycylglycine \xrightarrow{GGT} Glu-gly-gly + 5-Amino-2 - nitrobenzoate

The absorbance of this rate reaction is measured at 405/505 nm. The production is directly proportional to the GGT activity in the sample.

[Principle of Operation]

Refer to the Celercare V or the Pointcare V chemistry analyzer Operator's Manual, for the Principles and Limitations of the Procedure.

[Description of Reagents]

Each Equine Profile (15+4) contains lyophilized test-specific reagent beads. A lyophilized blank



reagent bead includes in each disc for a judgment of error 0209.

Type B is the reagent disc with diluent container.

Type A is the reagent disc without diluent container.

Calibration information is included in barcode code. Please check it on the label.

The component of each Equine Profile (15+4) is as follows (after redissolution):

Component	Quantity
Total protein assay reagent	13.5 μL
Albumin assay reagent	13.5 μL
Total Bilirubin assay reagent	13.5 μL
Urea assay reagent	13.5 μL
Creatinine assay reagent	13.5 μL
Glucose assay reagent	6.6 µL
Creatine kinase assay reagent	13.5 μL
Alkaline Phosphatase assay reagent	13.5 μL
Gamma Glutamyltransferase assay reagent	13.5 μL
Potassium assay reagent	13.5 μL
Sodiumassay reagent	13.5 μL
Calcium assay reagent	9.7 μL
Lactate dehydrogenase assay reagent	13.5 μL
Aspartate Aminotransferase assay reagent	13.5 μL
Total Carbon dioxide assay reagent	6.6 µL
Stabilizer	Appropriate amount

[Storage]

Store reagent discs in their sealed pouches at 2-8°C (36-46°F). Do not expose opened or unopened discs to direct sunlight or temperatures above 32°C (90°F). Reagent discs may be used until the expiration date included on the package. The expiration date is also encoded in the unique code printed on the sealing pouch. An error message will appear on the Celercare V or the Pointcare V chemistry analyzer display if the reagents have expired.

A torn or otherwise damaged pouch may allow moisture to reach the unused disc and adversely affect reagent performance. Do not use a disc from a damaged pouch.

Sample Requirements

Sample collection techniques are described in the "Sample requirement" section of the Celercare V or the Pointcare V chemistry analyzer Operator's Manual.



The required sample usage is $100 \,\mu\text{L}$ of lithium heparin plasma, serum or quality controls. Please add diluent when using Type A. The required diluent usage is $430 \,\mu\text{L}$ of sterilized water for injection.

At the same time, it is necessary to carry out the test within 60 minutes.

The glucose concentration is affected by the patient's feeding time and the storage environment after the sample is collected. In order to accurately measure glucose, a sample of the patient should be taken after at least 12 hours of fasting. For uncentrifuged samples stored at room temperature, the glucose concentration is reduced by about 5-12 mg / dL in 1 hour.

Light may cause total bilirubin to decompose, causing deviations in the test results.

Use only lithium heparin evacuated specimen collection tubes for plasma samples.

The test was started within 10 minutes after transferring the sample to the reagent disc.

【Interfering Substances】

Studies on known drugs or chemicals have found that when the interfering substances contained in the sample exceed the contents in the table below, the final test results are affected.

			Interferi	ng substances c	oncentration	(≤)		
A 1	Bilirubin	Intralipid	Hemoglobin	Vitamin C	Pyruvate	NH ₄ Cl	Creatine	Mg^{2+}
Analyte	mg/dL	mg/dL	mg/dL	mg/dL	mmol/L	mmol/L	μmol/L	mmol/L
TP	25	1050	200					
ALB	40	600	1000					
AST	40	600	50	25	1			
GGT	40	1050	200					
BUN	25	600	1000			1		
ALP	40	1050	400					
K^+	16	150	50	75				
Na^+	10	150	50	75				
Ca	180	210	200	75				3
LDH	40	1000	50	100				
TBIL		1050	1000	75				
CRE	40	1050	500	25			600	
GLU	40	600	1000	50				
CK	40	1000	400	100				
tCO_2	45	525	250	75				

[Procedure]

■ Materials Provided

Equine Profile (15+4)

Celercare V or Pointcare V chemistry analyzer

Please add diluent into the diluent port when using Type A (sterilized water for injection); please tear off the aluminum strip before using for Type B.

Transfer pipettes (fixed volume 100 μL for sample and 430 μL for diluent) and tips.

■ Test Procedure

The complete sample collection and step-by-step operating procedures are detailed in the Celercare V or the Pointcare V chemistry analyzer Operator's Manual.



■ Calibration

Each batch of reagent is calibrated using Randox standard serum to obtain the disc-specific calibration parameters before shipment.

The calibration parameters stored in the two-dimentional code printed on the sealed pouch are provided to analyzer at the time of scanning the code.

Refer to the Celercare V or the Pointcare V chemistry analyzer Operator's Manual for the specific information.

■ Quality Control

Refer to Operator's Manual of the Celercare V or the Pointcare V chemistry analyzer. Performance of the Celercare V or the Pointcare V chemistry analyzer can be verified by running controls. For a list of approved quality control materials with acceptance ranges.

If control results are out of range, repeat one time. If still out of range, call MNCHIP customer service or local distributers for technical support. Do not report the results if controls are outside their labeled limits.

Results

The Celercare V or the Pointcare V chemistry analyzer automatically calculates and prints the analyte concentrations in the sample. Details of the endpoint and rate reaction calculations are found in the Celercare V or the Pointcare V chemistry analyzer Operator's Manual.

[Normal Reference Ranges]

These ranges are provided as a guideline only. It is recommended that your office or institution establish normal ranges for your particular patient population.

Analyte	SI Units	Common Units
	Equine: 57 ~ 80g/L;	Equine: 5.7 ~ 8.0g/dL;
TP	Dog: 52 ~ 82g/L;	Dog: $5.2 \sim 8.2 \text{g/dL}$;
	Cat: 54 ~ 89g/L	Cat: 5.4 ~ 8.9g/dL
	Equine: 22 ~ 37g/L;	Equine: $2.2 \sim 3.7 \text{ g/dL}$;
ALB	Dog: 22 ~ 44g/L;	Dog: $2.2 \sim 4.4 \text{ g/dL}$;
	Cat: 22 ~ 45g/L	Cat: $2.2 \sim 4.5 \text{ g/dL}$
	Equine: $9 \sim 39 \mu mol/L$	Equine: 0.5~ 2.3 mg/dL
TBIL	Dog: $2 \sim 15 \mu \text{mol/L}$;	Dog: 0.1 ~ 0.9mg/dL;
	Cat:2 $\sim 15 \mu \text{mol/L}$	$Cat: 0.1 \sim 0.9 mg/dL$
	Equine: 175~ 340U/L;	Equine: 175 ~ 340U/L;
AST	Dog: 8.9 ~ 55U/L;	Dog: 8.9 ~ 55U/L;
	Cat: 9.2 ~ 60U/L	Cat: 9.2 ~ 60U/L
	Equine:5~24U/L;	Equine:5~24U/L;
GGT	Dog:0 ~ 7U/L;	Dog:0 ~ 7U/L;
	Cat: 0 ~ 2U/L	Cat: 0 ~ 2U/L
	Equine: 50 ~ 170U/L;	Equine: 50 ~ 170U/L;
ALP	Dog: 20 ~ 150U/L;	Dog: 20 ~ 150U/L;
	Cat:10 ~ 90U/L	Cat:10 ~ 90U/L



	Equine: 120 ~ 470U/L;	Equine: 120 ~ 470U/L;
CK	Dog: 20 ~ 200U/L;	Dog: 20 ~ 200U/L;
	Cat: 50 ~ 450U/L	Cat: 50 ~ 450U/L
	Equine: $3.6 \sim 6.1 \text{mmol/L}$	Equine:65 ~ 110mg/dL
GLU	Dog:3.89 ~ 7.95mmol/L	$Dog:70 \sim 143 mg/dL$
	Cat:4.11 ~ 8.84mmol/L	Cat:74 ~ 159mg/dL
	Equine:53 ~194 \(\mu \) mol/L	Equine: $0.6 \sim 2.19$ mg/dL;
CRE	Dog: 27 ~ 149μmol/L;	Dog: $0.3 \sim 1.7 \text{mg/dL}$;
	Cat:27 ~ 223 µmol/L	$Cat: 0.3 \sim 2.5 mg/dL$
	Equine:2.5 ~ 8.9mmol/L	Equine: $7 \sim 25 \mathrm{mg/dL}$
BUN	$Dog:2.5 \sim 11.5 mmol/L$	$Dog:7 \sim 32mg/dL$
	Cat:3.6 ~ 15.5mmol/L	$Cat:10 \sim 43 mg/dL$
	Equine: 20 ~ 33mmol/L;	Equine: 20 ~ 33mmol/L;
tCO_2	Dog: 12 ~ 27mmol/L;	Dog: 12 ~ 27mmol/L;
	Cat:15 \sim 24mmol/L	Cat:15 ~ 24mmol/L
	Equine: $2.9 \sim 3.6 \text{mmol/L}$;	Equine:11.5 ~ 14.2mg/dL;
Ca	Dog: 1.98 ~ 2.95mmol/L;	Dog: $7.9 \sim 11.8 \text{mg/dL}$;
	Cat: 1.95 ~ 2.95mmol/L	Cat: 7.8 ~ 11.8mg/dL
	Equine: $2.5 \sim 5.2 \text{mmol/L}$;	Equine: $2.5 \sim 5.2 \text{mmol/L}$;
K^+	Dog: $3.7 \sim 5.8$ mmol/L;	Dog:3.7 ~ 5.8mmol/L;
	Cat: 3.7 ~ 5.8mmol/L	Cat: 3.7 ~ 5.8mmol/L
	Equine: 126 ~ 146mmol/L;	Equine: 126 ~ 146mmol/L;
Na^+	Dog:138 ~ 160mmol/L;	Dog:138 ~ 160mmol/L;
	Cat: 142 ~ 164mmol/L	Cat: 142 ~ 164mmol/L
	Equine: 250 ~ 2100 U/L;	Equine: 250 ~ 2100 U/L;
LDH	Dog:40 ~ 400U/L;	Dog:40 ~ 400U/L;
	Cat: 0 ~ 800U/L	Cat: 0 ~ 800U/L

【Interpretation of Results】

Physiological interferents (hemolysis, icterus and lipemia) cause changes in the reported concentrations of some analytes. The sample indices are printed on the bottom of each printout to inform the operator about the abnormal sample. The operator should avoid sample hemolysis caused by irregular blood collection

The Celercare V or the Pointcare V chemistry analyzer suppresses any results that are affected by >10% interference from hemolysis, lipemia or icterus. "HEM", "LIP", or "ICT" respectively, is printed on the printout in place of the result.

For the same sample, the potassium result of using anticoagulant plasma is 0.2 - 0.5 mmol/L lower than those using serum. The potassium assay is a coupled pyruvate kinase (PK)/lactate dehydrogenase (LDH) assay. Therefore, in cases of extreme muscle trauma or highly elevated levels of creatine kinase (CK), The Celercare V or the Pointcare V chemistry analyzer may report a falsely elevated potassium (K $^+$) value. In such cases, unexpected high potassium recoveries need to be confirmed utilizing a different methodology.

Any result for a particular test that exceeds the assay range should be analyzed by another approved test



method or sent to a referral laboratory. Do not dilute the sample and run it again on the Celercare V or the Pointcare V chemistry analyzer.

【Limitations of Procedure】

The Equine Profile (15+4) should be used with the Celercare V or the Pointcare V chemistry analyzer, and is just used for in vitro diagnosis (IVD).

As with any diagnostic test procedure, all other test procedures including the clinical status of the patient, should be considered prior to final diagnosis.

[Performance Characteristics]

Accuracy

Analyta	The relative deviation or absolute deviation should meet the following
Analyte	requirements
TP	B% ≤ 5.0%
ALB	$B\% \leq 6.0\%$
ALP	$B\% \le 10.0\%$
TBIL	$B\% \leq 10.0\%$
CRE	$B\% \le 10.0\%$
BUN	$B\% \le 15.0\%$
GLU	$B\% \leq 20.0\%$
CK	$B\% \le 10.0\%$
AST	$B\% \le 15.0\%$
LDH	$B\% \leq 10.0\%$
\mathbf{K}^{+}	$B\% \le 15.0\%$
Na^+	$B\% \le 15.0\%$
Ca	$B\% \leq 5.0\%$
GGT	$B\% \le 15.0\%$
tCO ₂	$B\% \leq 10.0\%$

Batch precision

Analyte	Coefficient of variation (≤ *)
TP	2.0%
ALB	2.0%
ALP	5.0%
TBIL	5.0%
CRE	5.0%
BUN	5.0%
GLU	5.0%
CK	5.0%
AST	5.0%
LDH	5.0%
K ⁺	5.0%



Na ⁺	5.0%
Ca	3.0%
GGT	5.0%
tCO_2	5.0%

Inter batch precision

Analyte	Relative Range (≤ *)
TP	5.0%
ALB	5.0%
ALP	10.0%
TBIL	10.0%
CRE	10.0%
BUN	10.0%
GLU	10.0%
CK	10.0%
AST	10.0%
LDH	10.0%
$\mathbf{K}^{\scriptscriptstyle +}$	10.0%
Na^+	10.0%
Ca	5.0%
GGT	10.0%
tCO ₂	10.0%

Dynamic Ranges

Analyte	Dynamic Ranges
TP	20 ~100g/L
ALB	10~60g/L
ALP	5 ~ 2000U/L
TBIL	2~800μmol/L
CRE	$20{\sim}2000\mu mol/L$
BUN	0.9 ~35.7mmol/L
GLU	1 ~35mmol/L
CK	5 ~ 3000U/L
AST	5 ~ 1600U/L
GGT	5 ~ 1500U/L
LDH	25 ~ 3000U/L
K^+	1 ~ 8 mmol/L
Na ⁺	90 ~ 170mmol/L



Ca	0.5 ~ 4mmol/L
tCO_2	10 ~ 35mmol/L

Notes

Used reagent discs contain animal body fluids. Follow good laboratory safety practices when handling and disposing of used discs. See the Celercare V or the Pointcare V chemistry analyzer Operator's Manual for instructions on cleaning biohazardous spills.

The reagent discs are plastic and may crack or chip if dropped. Never use a dropped disc as it may spray biohazardous material throughout the interior of the analyzer.

Reagent beads may contain acids or caustic substances. The operator does not come into contact with the reagent beads when following the recommended procedures. The operator should avoid ingestion, skin contact, or inhalation of the reagent beads.

The diluent can be selected from purified water having a conductivity(measured at 25°C) greater than $10~M\Omega/cm$, we recommend using the sterilized water for injection to reduce discrepancies or errors in test results due to the water, and it should be prevented from being exposed to the air for a long time after opening.

[Symbols Used in Labelling]

Symbol	Explanation
Veterinary	Veterinary use only
	Manufacturer
EC REP	Authorized representative in the European Community
\square	Use-by date
LOT	Batch code
~~ <u> </u>	Date of manufacture
[]i	Consult instructions for use
2°C 18°C	Limit of temperature
((())	Do not re-use

[Manufacturer]



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EC REP

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